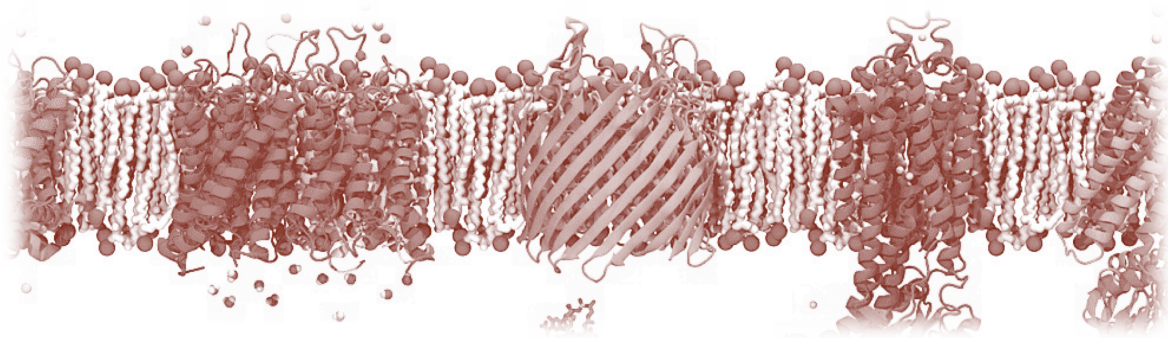
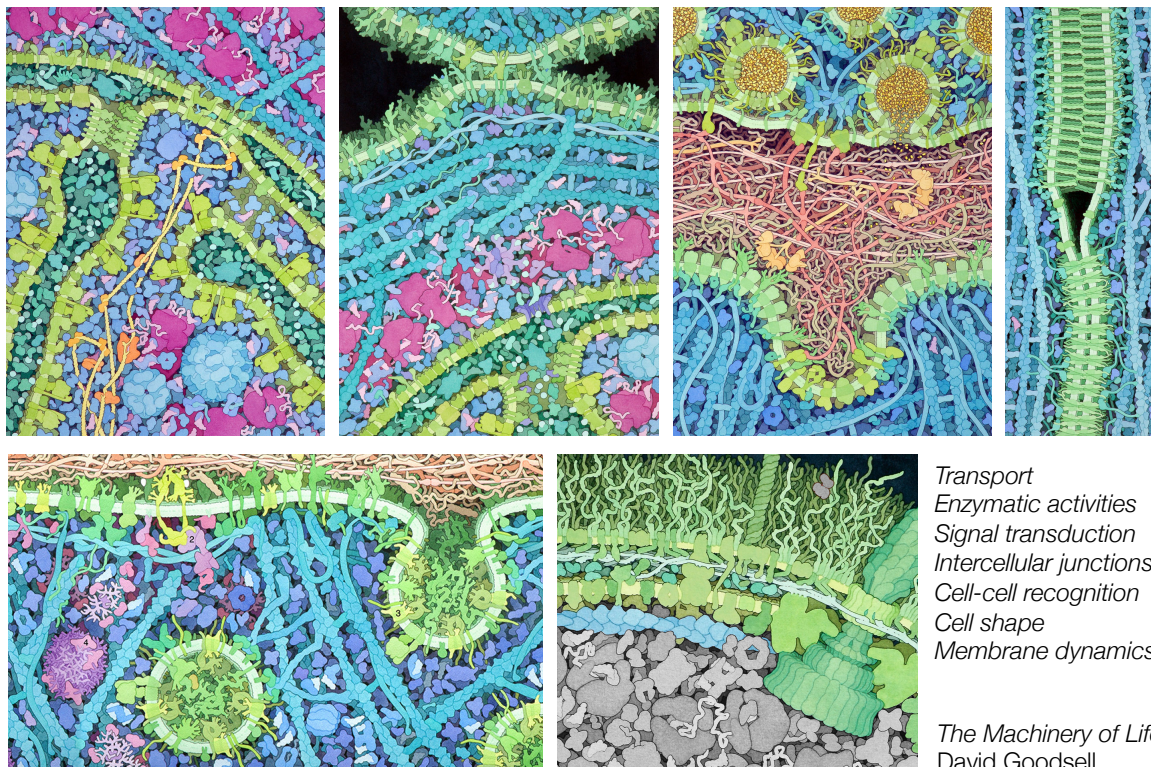


Membrane Proteins: Structural & Functional Challenges



Susana Andrade
Institute of Biochemistry
University Freiburg

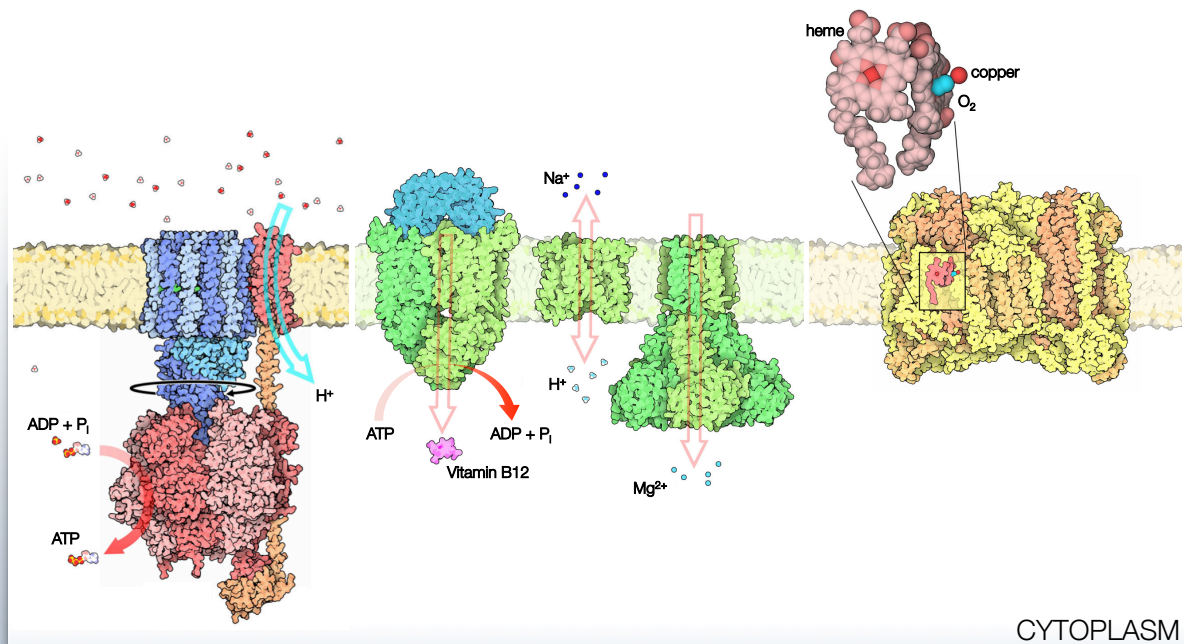
Diverse roles and structures



Transport
Enzymatic activities
Signal transduction
Intercellular junctions
Cell-cell recognition
Cell shape
Membrane dynamics

The Machinery of Life,
David Goodsell

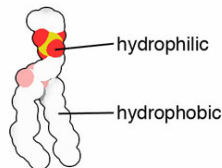
Zooming in



The membrane environment

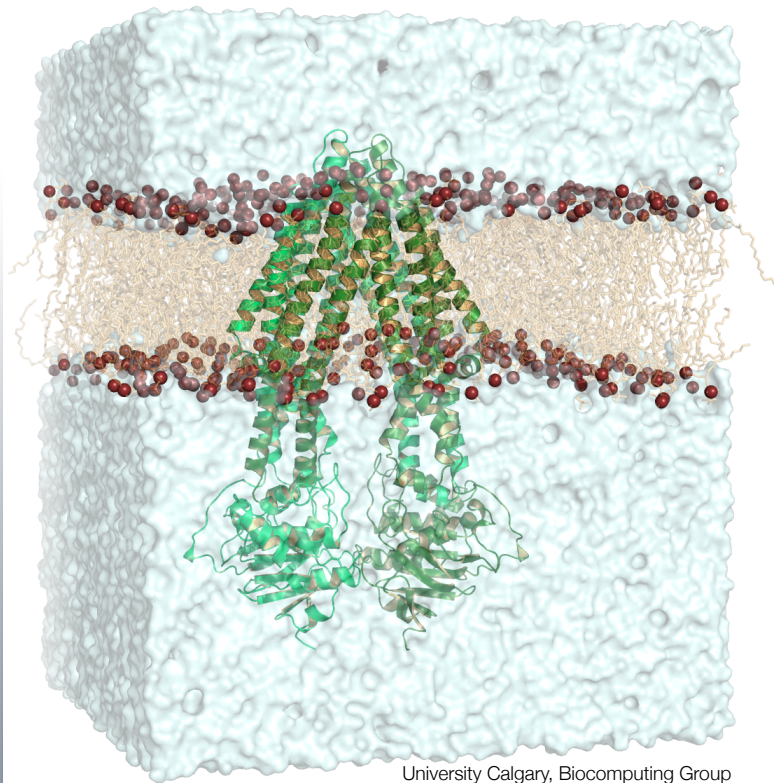
Composition

- **Glycerophospholipids**
 - Phosphatidic acid
 - Phosphatidylinositol
 - Phosphatidylserine
 - Phosphatidylcholine
 - Phosphatidylethanolamine
 - Phosphatidylglycerol
 - Diphosphatidylglycerol
 - Variable size and saturation of the aliphatic chains
 - (...)
- **Glyceroglycolipids**
 - (...)
- **Sphingophospholipids**
 - (...)
- **Sphingoglycolipids**
 - (...)
- **Sterols**
 - (...)
- **Others**
 - Dolichols
 - (...)

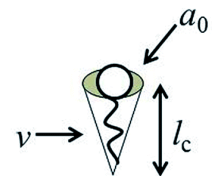


Function

- **Barriers**
 - Create variable compartments
- **Support (for membrane proteins)**
 - Selective barrier, membrane curvature, inner/outer leaflet properties
- **Modulate protein function**
 - Act as cofactors
 - Transverse forces (hydrophobic mismatch)
 - Lateral forces
 - Lipid rafts
- **Signaling**
 - PI, DAG, ceramide, PS, (...)
- **Reservoir of lipids for the cell**
 - Energy, Signal molecules/precursors (...)
- **Others**
 - (...)

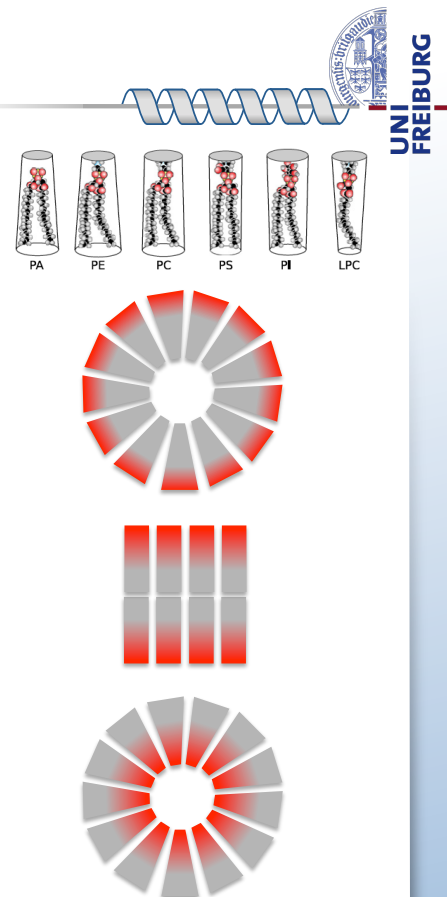


University Calgary, Biocomputing Group

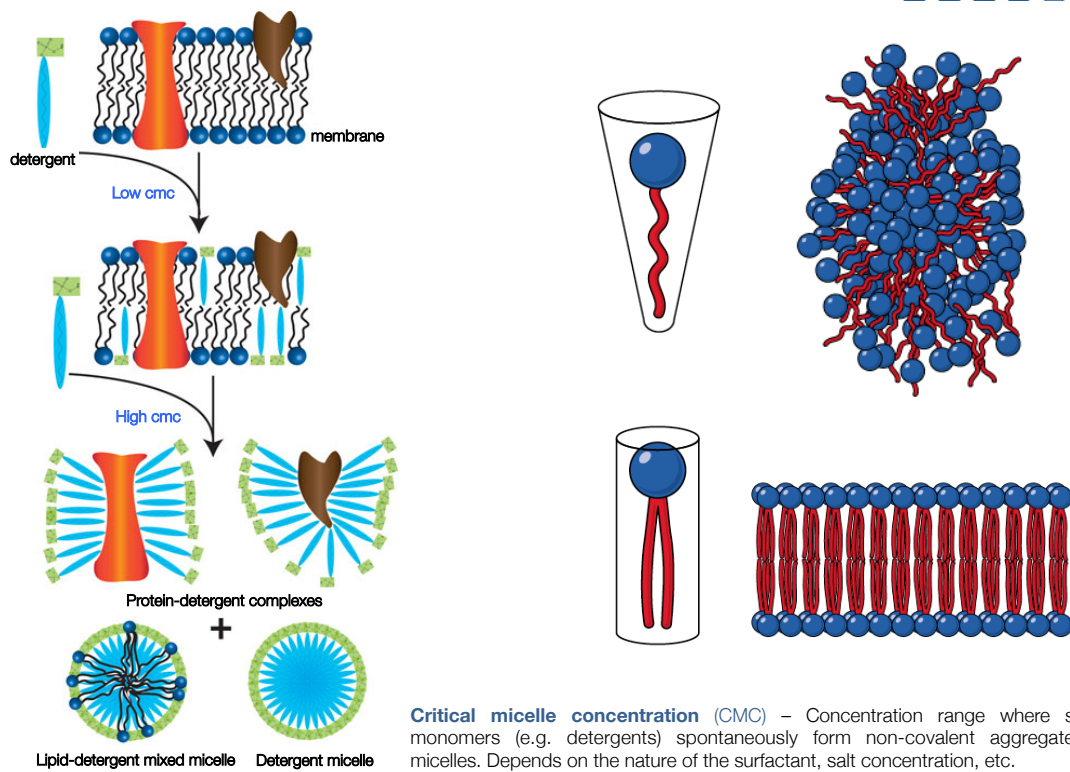


$$CPP = v/a_0 l_c$$

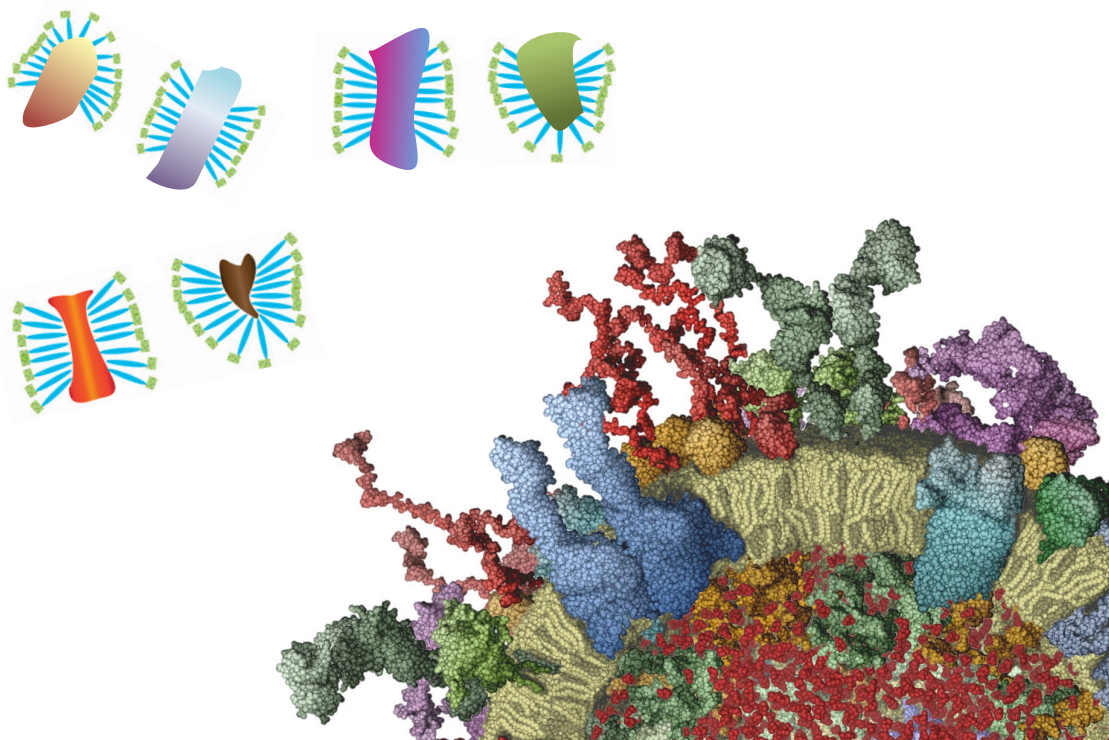
Critical Packing Parameter ($v/a_0 l_c$)	Critical Packing Shape	Structures Formed
$< 1/3$	Cone	Spherical micelles
$1/3 - 1/2$	Truncated cone	Cylindrical micelles
$1/2 - 1$	Truncated cone	Flexible bilayers, vesicles
~ 1	Cylinder	Planar bilayers
> 1	Inverted truncated cone or wedge	Inverted micelles



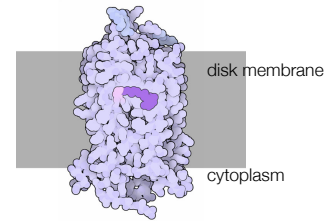
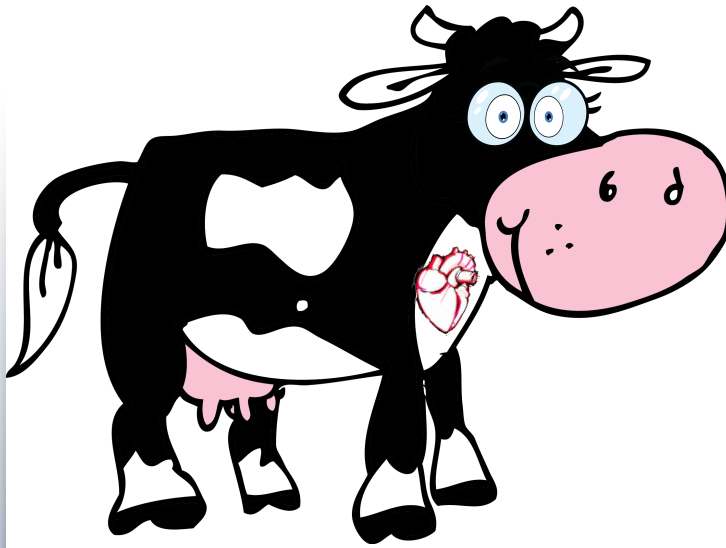
Solubilizing membrane proteins



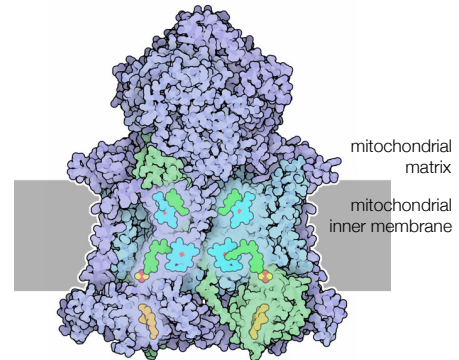
Purifying membrane proteins



Purifying membrane proteins



Rhodopsin



Cytochrome bc1

Producing (membrane) proteins

To produce the protein encoded by a piece of cloned DNA, expression plasmids/vectors are required:

- **Promoter** (*inducible* vs *constitutive*) : allows regulating amount and time of protein expression
- **Antibiotic**: allows selecting cells carrying the plasmid
- **Ori**: origin of replication/replicon is the place where DNA replication begins, enabling a plasmid to reproduce itself
- **Multiple cloning region**: short segment of DNA with multiple restriction sites. It allows inserting a gene at a precise position.
- ...

❖ Common DNA sources and delivery mechanisms are **plasmids**, **viruses** (e.g. baculovirus, retrovirus, adenovirus), **artificial chromosomes** and **bacteriophage** (such as lambda).

Producing (membrane) proteins



Cell-based expression system:

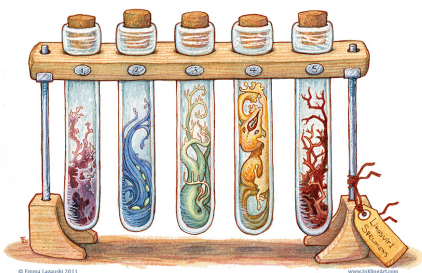
- The plasmid is placed *inside a cell*

Common hosts are **bacteria** (e.g. *E. coli*, *B. subtilis*), **yeast** (e.g. *S. cerevisiae*, *P. pastoris*), **eukaryotic cell lines** (HeLa, HEK, (...)).



The best expression system depends on the characteristics of the protein to produce:

- Bacterial – can produce large amounts of protein. Post-translational modifications and folding (inclusion bodies) can be a problem.
- S. cerevisiae* – when significant post-translational modifications are required.
- Insect or mammalian cell lines – for human-like splicing of mRNA. Glycosylation, (...).
- (...)



Cell-free expression system:

- in vitro*, with purified RNA polymerase, ribosomes, tRNA and ribonucleotides.

Producing (membrane) proteins



(...)

RNA polymerase will produce mRNA.

Ribosomes translate mRNA into a **protein**.

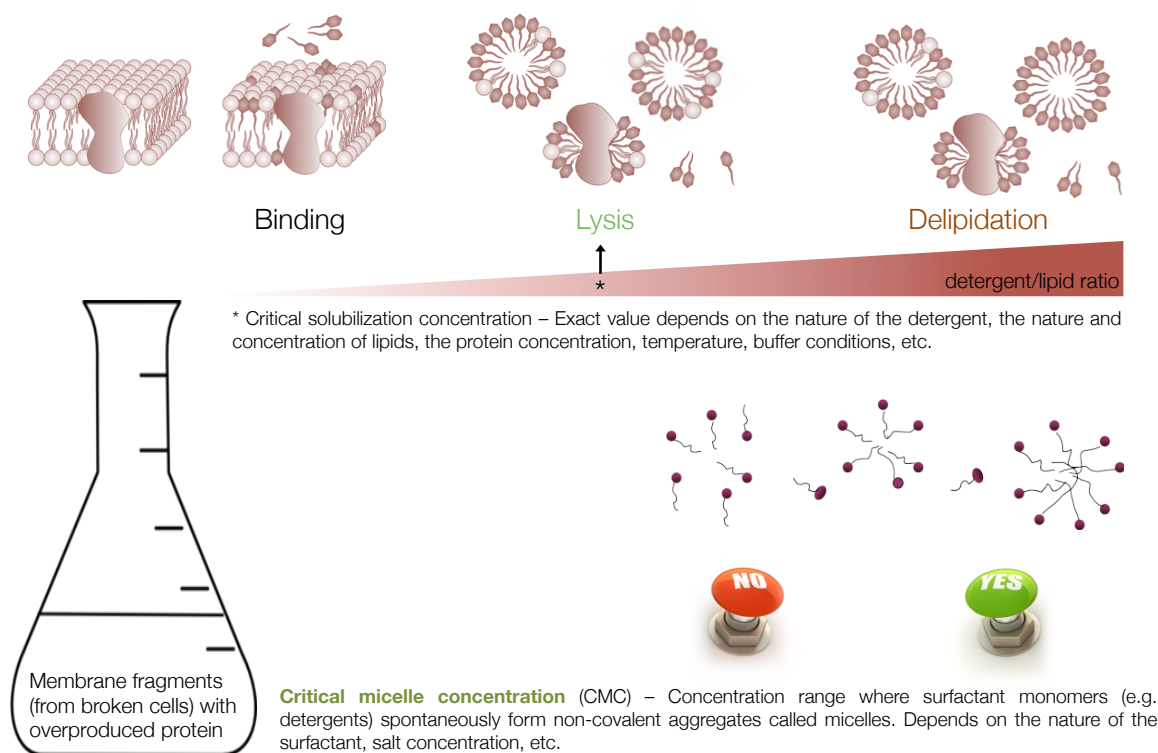
- Clone (homolog genes)
- Clone (plasmid/promoter type)
- Express (host cells, growth conditions)
- Protein targetting, yield
- Protein functionality
- Protein purification scheme



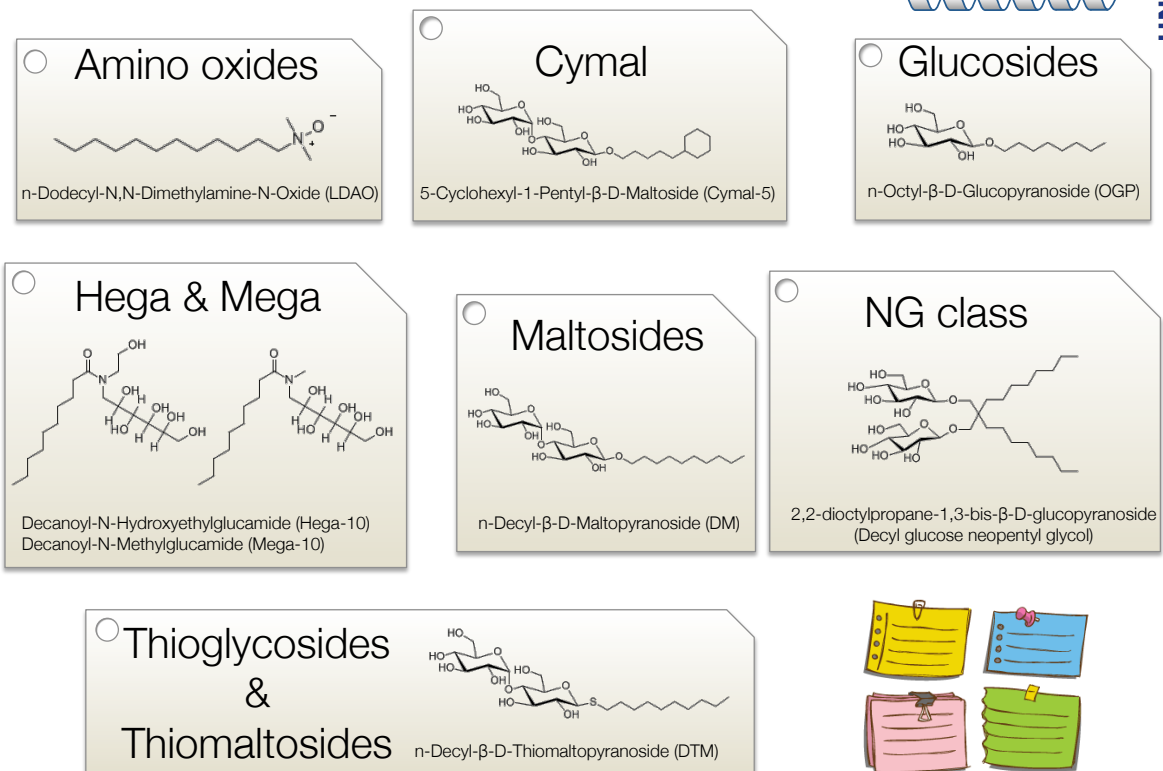
Epitope tags (portion of a molecule where antibody binds) can be added to help visualization by western blot or immunofluorescence.

Peptides can be added to increase solubility and detection (e.g. MBP, GFP)...

Purifying membrane proteins



The detergent variable

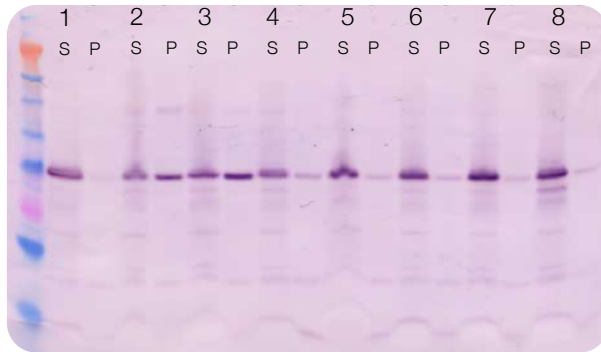


The detergent variable



Solubilization test:

e.g. Western blot* of solubilized (S) and un-solubilized (P) membranes



- 1: SDS
- 2: HEGA-10
- 3: C₈E₄
- 4: LDAO
- 5: DDM
- 6: Triton X-100
- 7: FOS-CHOLINE-12
- 8: OGP

Epitope tags (portion of a molecule where antibody binds) can be added to help visualization by western blot or immunofluorescence.

Peptides can be added to increase solubility and detection (e.g. MBP, GFP)...

*Rath et al (2007), PNAS, 106:1760

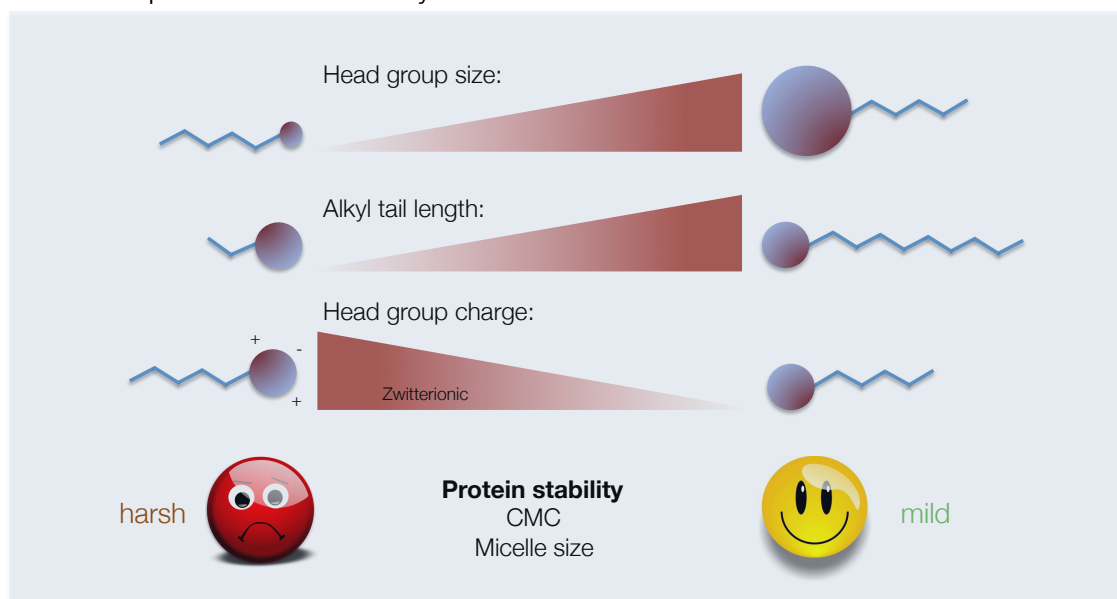
The detergent variable



1. Protein solubilization ✓

GOAL – Extract efficiently the target membrane protein and keep it stable in solution.

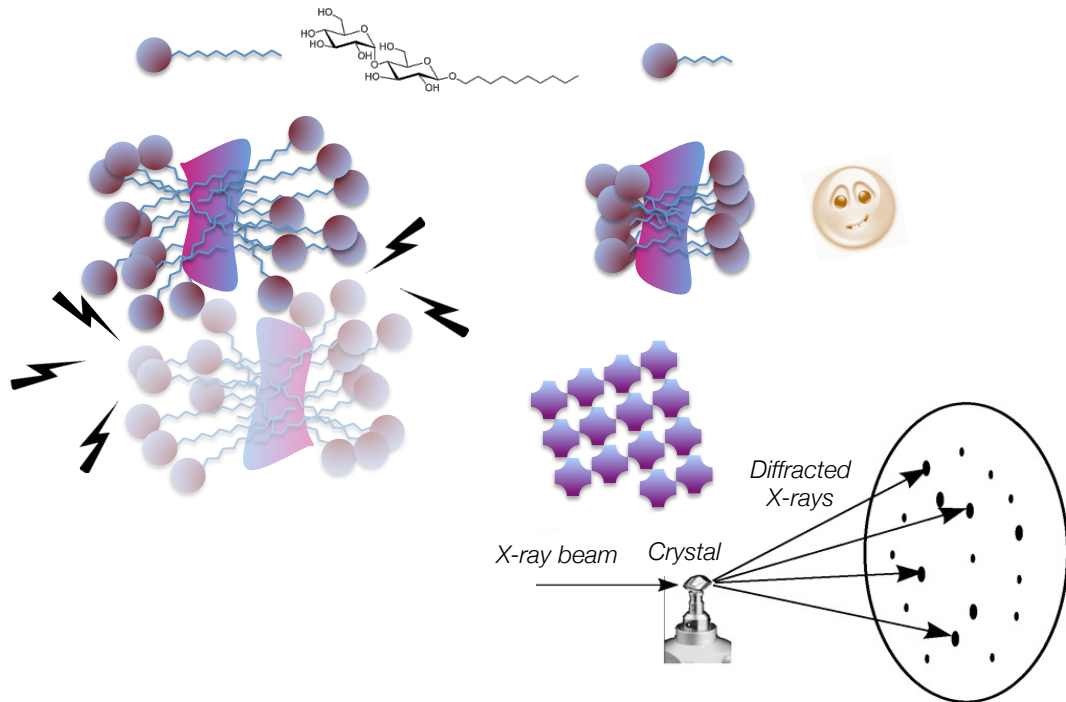
2. Protein purification & stability



The beauty and the beast

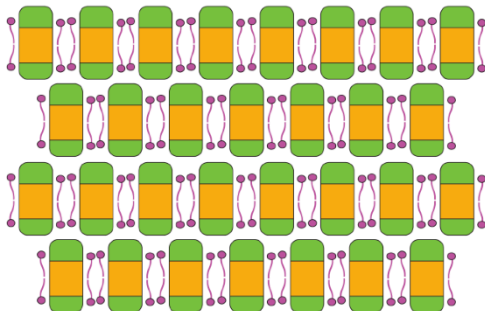
3. Protein characterization

- *X-ray crystallography*

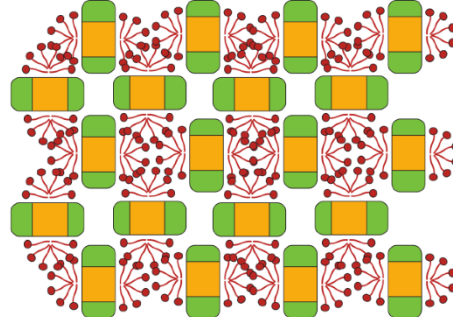


Types of membrane protein crystals

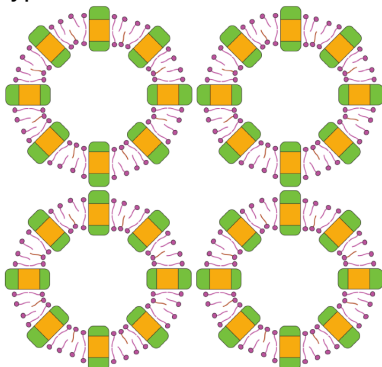
- Type I



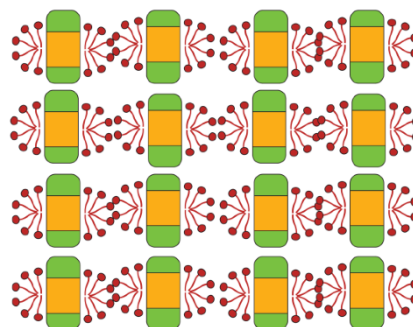
- Type II



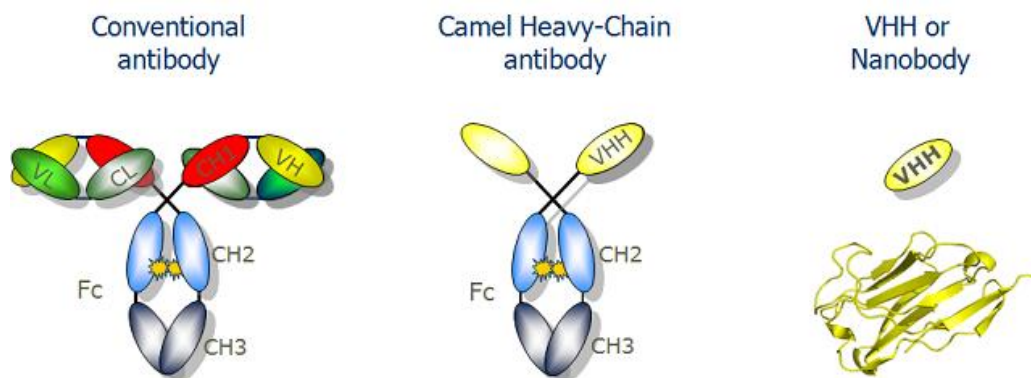
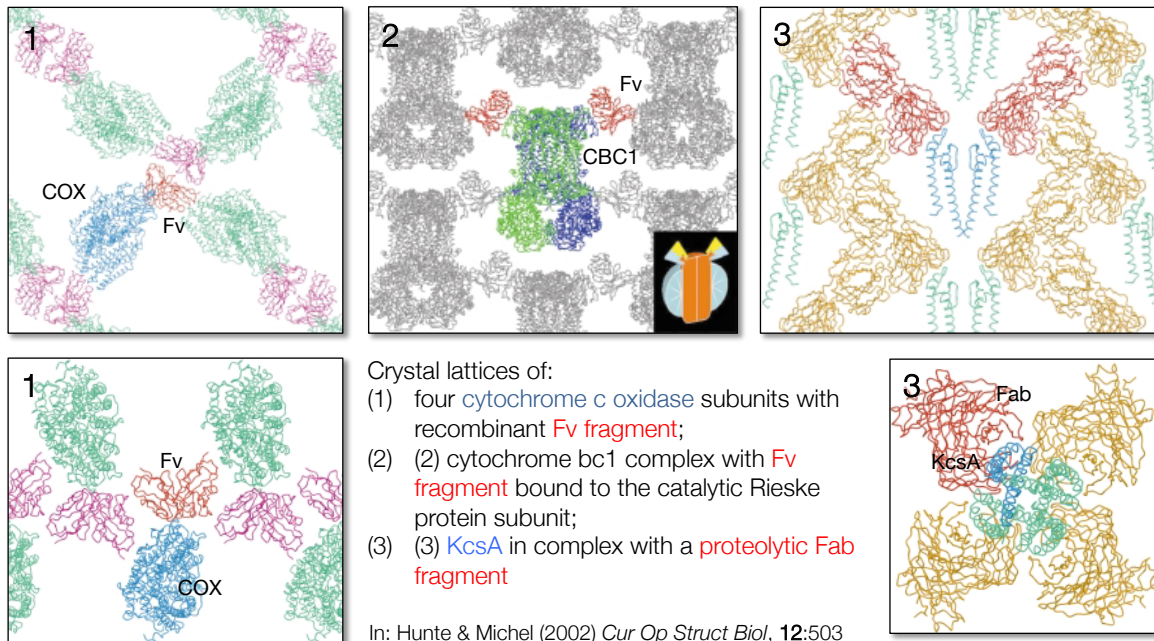
- Type III



- Type IV



Monovalent antibody fragments can be generated recombinantly as **Fv** (fragment variable ~28 kDa) or **Fab** (fragment antibody binding ~56 kDa) or by proteolytic cleavage.



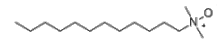
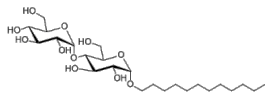
Recombinant Nanobodies are small (15kDa), monomeric, bind target with nM affinity, are stable, easy to manipulate and are well expressed in bacterial expression systems so that they are cheaper and easier to produce in all kind of formats than standard monoclonal antibodies.

Nanobodies often bind to epitopes that are less immunogenic for conventional antibodies, such as the active sites of enzymes.

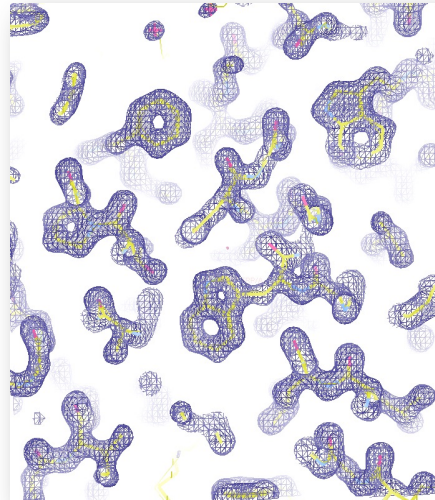
Due to their small size, they also target areas that are not accessible to standard antibodies.

Another advantage is that they generally bind conformational epitopes

High-resolution protein crystal structures

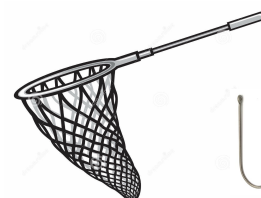
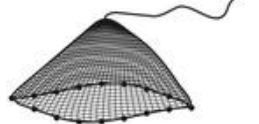
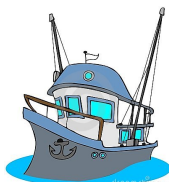


- Identify residues and elements
- Detect conformational changes
- Discuss molecular interactions
- Visualize pores/channels/cavities
- Reveal lipid/detergent binding sites
- Identify functional residues & locations (active site, selectivity filter, gating, ...)
- High level of disorder due to high flexibility



Electron density map of Af-Amt1 at 1.3 Å resolution.

Never give up





We have analyzed the local structure and dynamics of the prokaryotic voltage-dependent K⁺ channel (KvAP) at 0 millivolts, using site-directed spin labeling and electron paramagnetic resonance spectroscopy. We show that the α -segment is located at the protein/lipid interface, with the α -helix protected from the lipid environment. *Strand* separated into two short α -helices. *Angew. Chem. Int. Ed.* 2011, 50, 1302–1305

Lichi Shi, Izuru Kawamura, Kwang

Maofu Liao^{1*}, Erhu Cao^{2*}, David Julius² & Yifan Cheng¹

Transient receptor potential (TRP) channels are sensors for a wide range of cellular and environmental signals, but elucidating how these channels respond to physical and chemical stimuli has been hampered by a lack of detailed structural information. Here we exploit advances in cryo-electron microscopy to determine the structure of a mammalian TRP channel, TRPV1, at 3.5 Å resolution. TRPV1 is a homotrimeric ion channel with a large extracellular domain, a narrow selectivity filter, and a large intracellular domain. Like voltage-gated channels, TRPV1 exhibits four-fold symmetry around a central ion pathway formed by transmembrane segments 5–6 (S5–S6) and the intervening pore loop, which is flanked by S1–S4 voltage-sensor–like domains. TRPV1 has a wide extracellular “mouth” with a short selectivity filter. The conserved “TRP domain” intrinsically folds into a β -barrel structure, which is stabilized by a conserved disulfide bond. TRPV1 interacts with other proteins in the cell, including ankyrin, through its intracellular domain. These observations provide a structural blueprint for understanding unique aspects of TRP channel function.

For cells to function properly, membrane proteins must be able to diffuse within biological membranes. The functions of these proteins depend on their position within the membrane, and not only on their structure but also on their mobility. In this review, we discuss the role of membrane proteins in the function of the cell, and the importance of their mobility in the function of the cell. We discuss the role of membrane proteins in the function of the cell, and the importance of their mobility in the function of the cell. We discuss the role of membrane proteins in the function of the cell, and the importance of their mobility in the function of the cell.

Christiano Luis Pinto Oliveira,^[a] Daniel Erik Otzen,^[a, c, d] Marianne Glasius,^[a, b] Brian Stougaard Vad,^[a, c] Kell Kleiner Andersen,^[a, c] and Jan Skov Pedersen^[a, b]

ChemBioChem 2014, 15, 2113 – 2124

to determine the low-resolution structure of protein OMPA from *E. coli* solubilized with maltodextrin (DOM). We have

linked dimer as one and

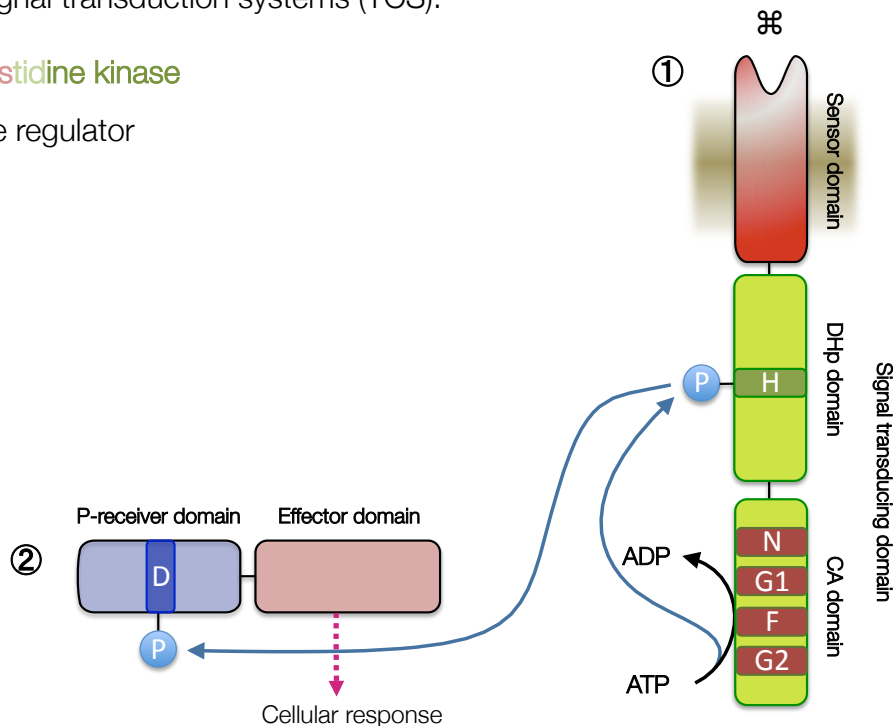
doi:10.1038/nature12822

We determine the low-resolution structure of protein OmpA from *E. coli* stabilized by methyl maltoide (DDM). We have studied transmembrane domain of OmpA—its associated dimers, and covalently linked dimer as the monomeric species of the full-length protein. We can successfully study the monomeric and covalently linked dimer as one and two natively folded proteins, respectively, whereas the noncovalently linked dimer presents a more complicated structure possibly due to its length problem. We have determined the structure of the full-length protein attached through a flexible linker to the transmembrane domain. This approach provides valuable information about membrane proteins as they are embedded in amphiphilic environment.

ChemBioChem 2014, 15, 2113–2124

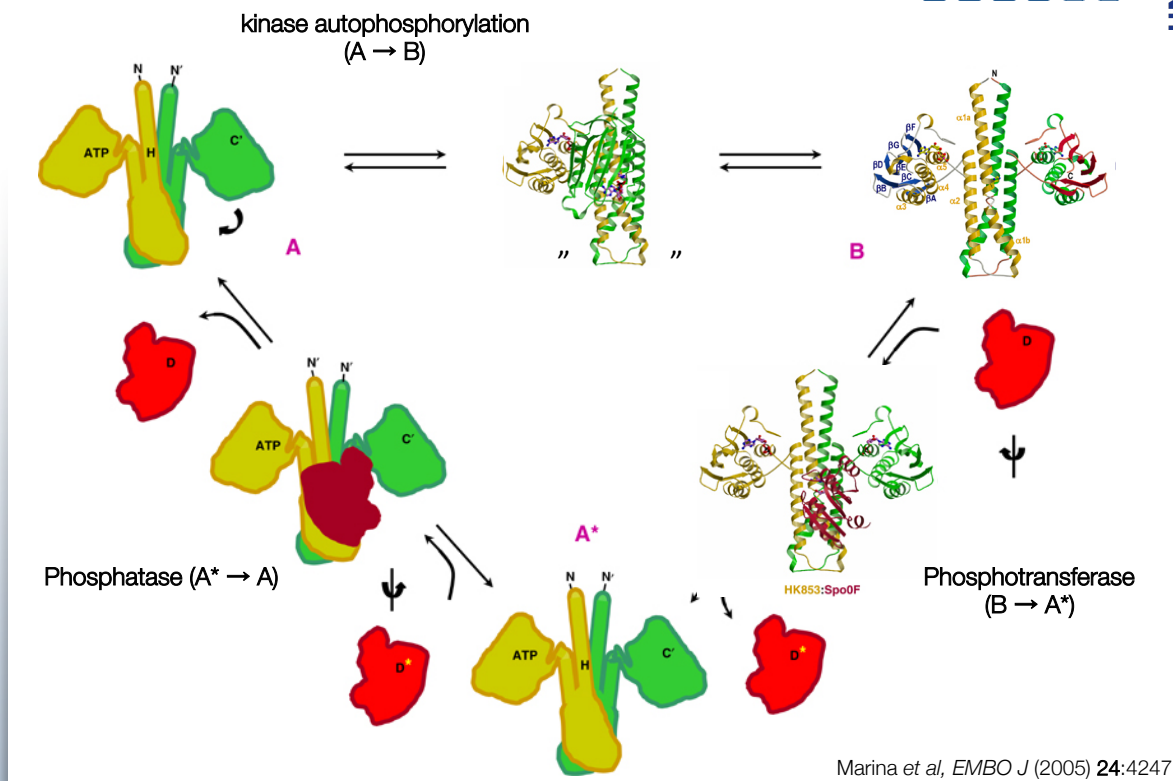


② - Response regulator



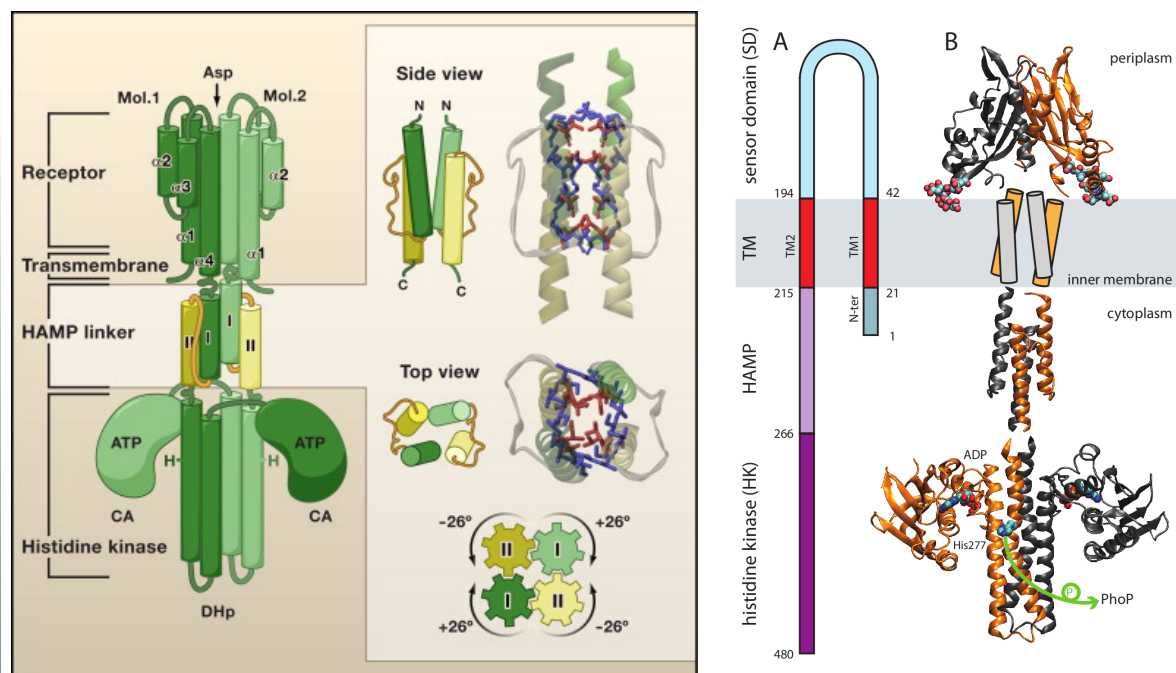
Cellular response

Structure-based scheme of the HK reactions

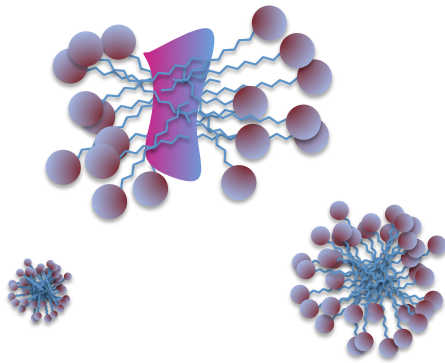


How is signal transmitted after reception?

E.g. – HAMP domains are typically associated with membrane domains and relay extracellular signals into intracellular responses. A unifying mechanism for HAMP domain signal transduction has yet to emerge, mainly due to lack of structural information.

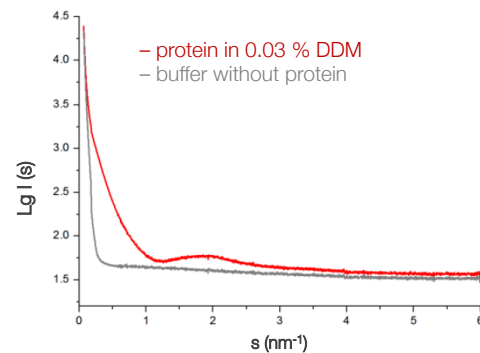
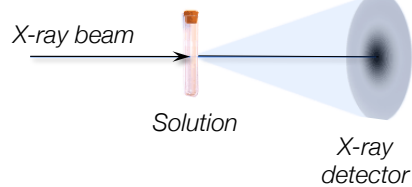


SAXS of the full-length sensor histidine kinase



Protein is pure, stable and reveals an homogeneous trimeric form in buffer containing 10 % glycerol and 0.03 % DDM.

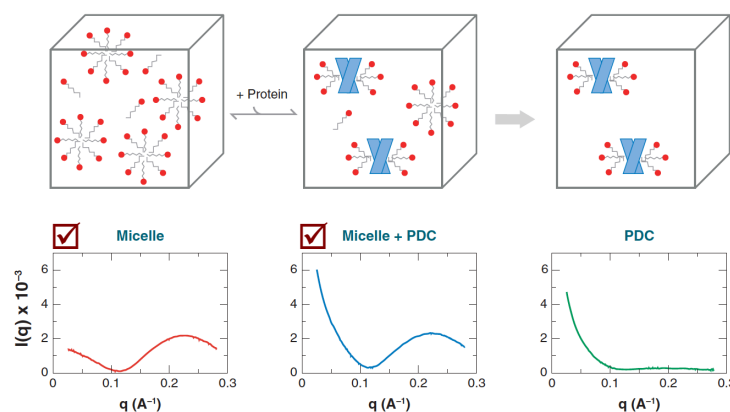
Dmitri Svergun, EMBL, Hamburg



The beauty and the beast



How to get rid of detergent backscattering from protein scattering?



Adapted from:
Lipfert & Doniach (2007)
Ann Rev Biophys Biomol Struct 36:307.



“Density matching” – Match the scattering density (contrast) of the solvent to that of the detergent..



“Subtracting micellar scattering” – Separate the contributions of the detergent micelles in the presence and absence of protein..



“Singular value decomposition” – Collect data at various protein:detergent ratios and apply a global fitting procedure..

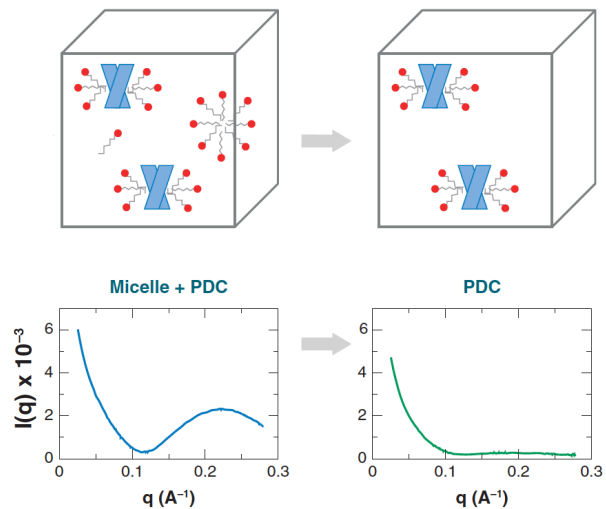
The beauty and the beast



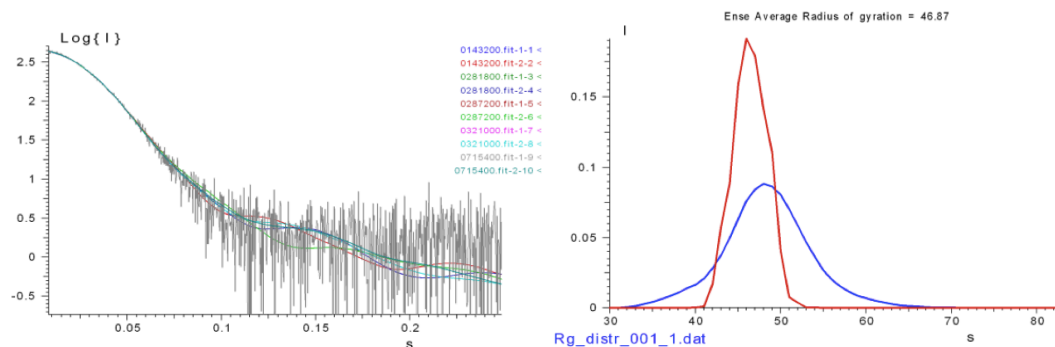
How to get rid of detergent backscattering from protein scattering?



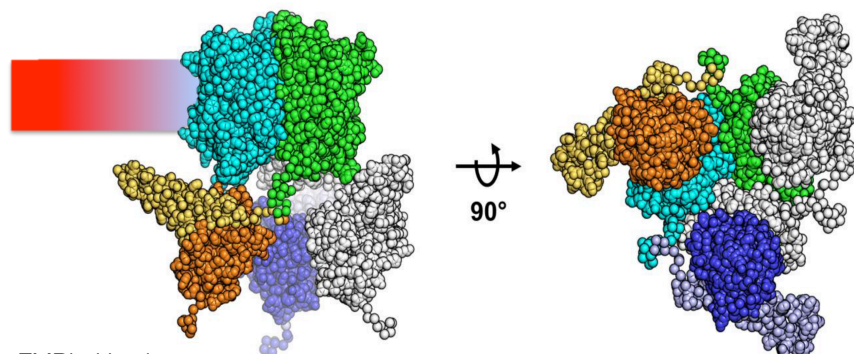
- **Dialysis** – Detergents with high CMCs are easily removed by dialysis. So that micelles disintegrate into monomers that easily pass through dialysis tubing over time.
- **Hydrophobic beads** – Detergents with low CMCs are typically removed by adsorption to hydrophobic beads (bio-beads) followed by filtration or centrifugation.
- **Chromatography** – Gel filtration can be used to separate detergent micelles from protein-detergent complexes and free protein based on size differences. Detergents can also be removed or exchanged by affinity chromatography.



SAXS of the full-length sensor histidine kinase



Model obtained using crystal structure models and the *ensemble optimization method* (EOM):

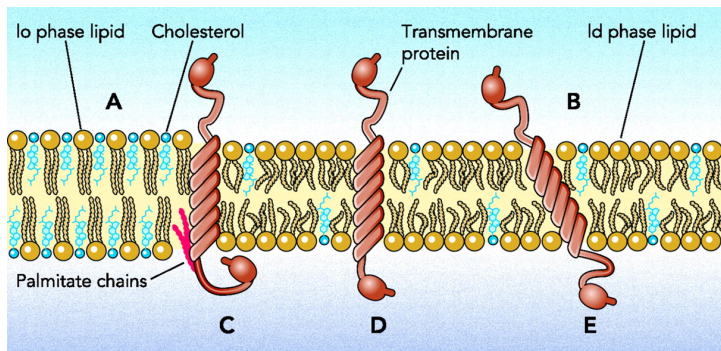
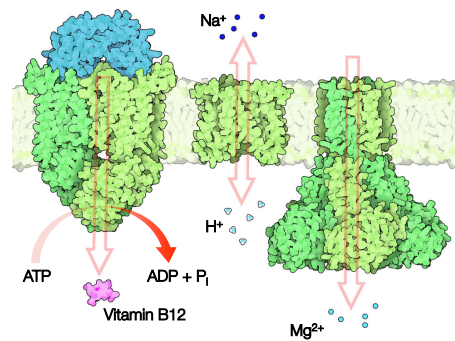


Dmitri Svergun, EMBL, Hamburg

The beauty and the beast (...)

The lipid environment can severely influence the protein

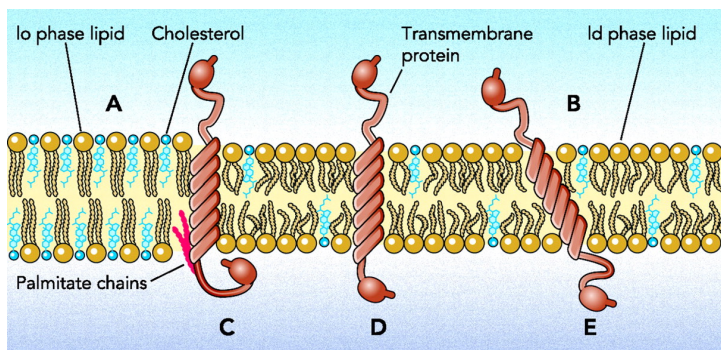
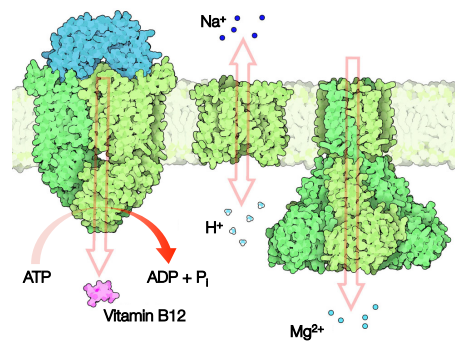
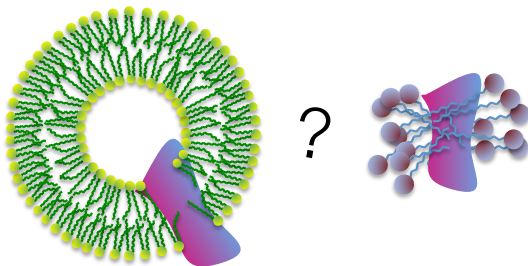
- fold
- activity



The beauty and the beast (...)

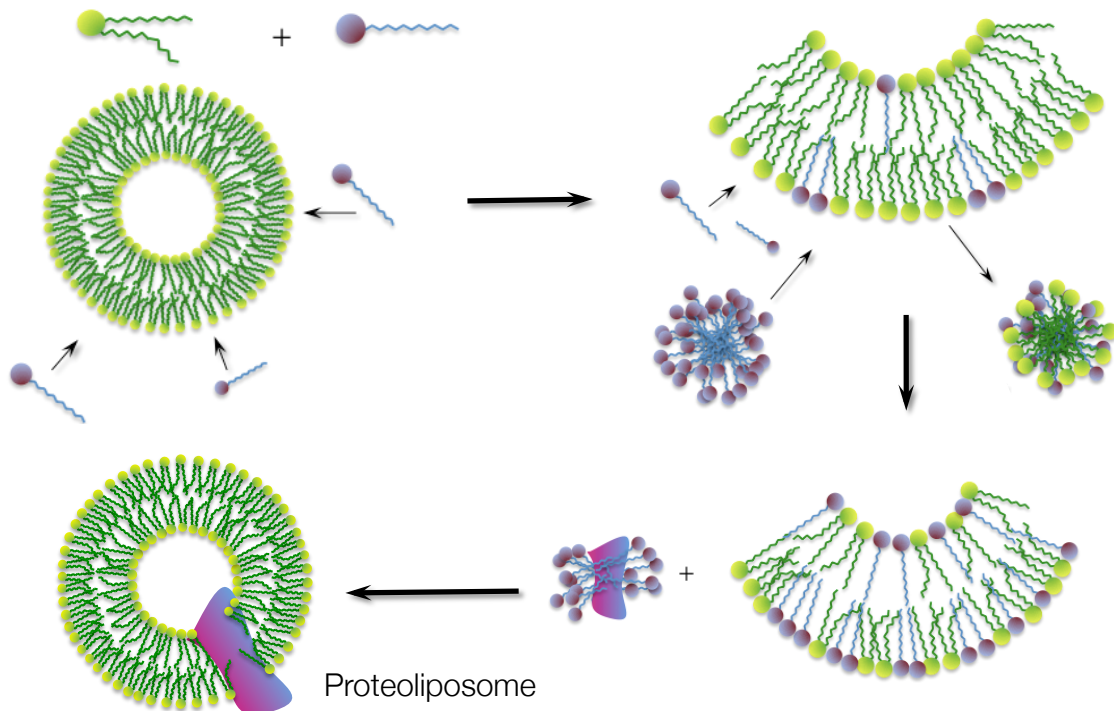
3. Protein characterization (cont.)

- *Functional assays*



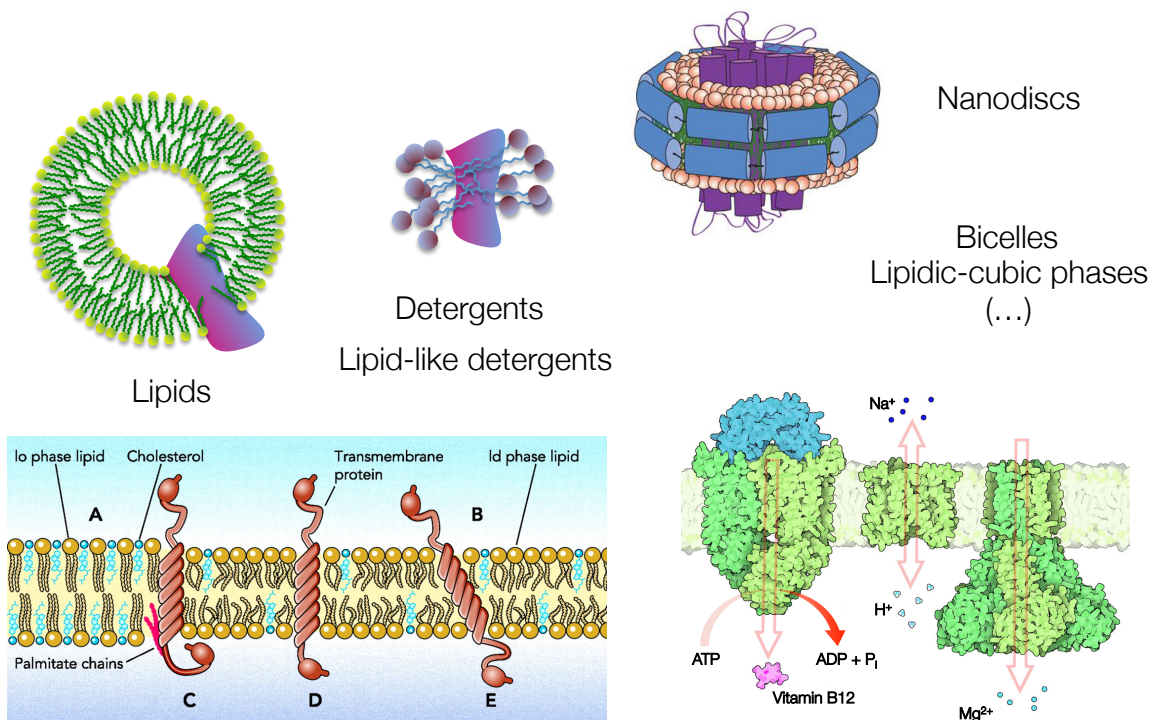
Reconstitution of membrane proteins

Liposome solubilization:



NEVER GIVE UP

3. Protein characterization



Be prepared and dare!

NEVER GIVE UP

